

# Mesoproterozoic carbon dioxide levels inferred from calcified cyanobacteria

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## ABSTRACT

Filamentous and shrub-like carbonate fabrics produced by *in vivo* cyanobacterial sheath calcification in stromatolites of the ca. 1200 Ma Society Cliffs Formation, Baffin and Bylot Islands, Arctic Canada, are 400 m.y. older than previously reported examples. *In vivo* sheath calcification is promoted by carbon dioxide concentrating mechanisms (CCMs) and is a direct ecophysiological link to atmospheric CO<sub>2</sub> concentration. CCMs are induced in present-day cyanobacteria under experimental conditions when pCO<sub>2</sub> is below ~0.36% (~10 times present atmospheric level, PAL). Society Cliffs calcified cyanobacteria consequently imply pCO<sub>2</sub> levels of <0.36% at ca. 1200 Ma. This inference is consistent with marine carbon isotope modeling that suggests pCO<sub>2</sub> of 7–10 PAL in the late Mesoproterozoic. Combined, petrographic, experimental, and modeling results therefore suggest that Mesoproterozoic pCO<sub>2</sub> concentrations were not substantially different from Phanerozoic values and were significantly less than previous estimates of up to 200 PAL. Assuming 10% lower solar luminosity in the late Mesoproterozoic, pCO<sub>2</sub> levels of 10 PAL or less require the presence of additional greenhouse gases for maintenance of an ice-free Earth. At 10 PAL pCO<sub>2</sub>, methane concentrations of 100–200 ppm would have been sufficient to sustain warm Earth surface conditions. The low atmospheric oxygen and limited marine sulfate concentrations required to sustain atmospheric methane provide additional support for sulfur isotope models that suggest protracted oxygenation of Earth's Proterozoic biosphere.

**Keywords:** Proterozoic, cyanobacteria, photosynthesis, calcification, carbon dioxide.

## INTRODUCTION

In the presence of a faint young sun, Earth's early atmosphere must have contained greenhouse gases sufficient to overcome lower temperatures driven by decreased solar luminosity (Sagan and Mullen, 1972). Traditionally, carbon dioxide and water vapor have been assumed to have been the principal greenhouse gases (Owen et al., 1979), and theoretical models require pCO<sub>2</sub> concentrations of 50–900 times present atmospheric level (PAL) 2.5 b.y. ago, and 10–400 PAL 1.5 b.y. ago, to avoid prolonged global glaciation (Kasting, 1987). Empirical estimates based on paleosols and weathering rinds, however, suggest pCO<sub>2</sub> levels in or below the lower part of this modeled range from the late Archean (Hessler et al., 2004) to the Mesoproterozoic (Rye et al., 1995; Sheldon, 2006). These relatively low estimates of pCO<sub>2</sub> imply the presence of additional greenhouse gases, such as methane, for maintenance of nonfreezing surface conditions (Pavlov et al., 2003; Kasting, 2005). Biological production and photochemical survival of methane would have been enhanced in low-oxygen environments of the Archean and early Proterozoic (Kasting et al., 1983; Goldblatt et al., 2006), and models suggest that methane values could readily have reached ~400 ppm (Pavlov et al., 2001). Furthermore, at high atmospheric concentrations, methane acts as an effective barrier to solar input (methane haze), which could trigger episodes of global cooling consistent with "snowball" glaciations in the early and late Proterozoic (Pavlov et al., 2001; Schrag et al., 2002; Pavlov et al., 2003).

Methane flux, however, is strongly dependent upon atmospheric pO<sub>2</sub> and marine sulfate concentration, both of which may have changed dramatically after ca. 2.2 Ga (Canfield, 1998; Farquhar et al., 2000). Because methane levels cannot currently be constrained from the geologic record, empirical measurements of pCO<sub>2</sub> remain critical to understanding the long-term evolution of Earth's atmosphere. Few estimates of Proterozoic pCO<sub>2</sub> levels have been based on fossils. Comparisons of carbon isotope composition of Mesoproterozoic (1.4 Ga) acritarchs with specific photosynthetic carbon-fixation pathways restrict pCO<sub>2</sub> levels only within a wide range, from >10 to 200 PAL (Kaufman and Xiao, 2003). Here, we use *in vivo* cyanobacterial calcification as a barometer for paleo-pCO<sub>2</sub> and infer late Mesoproterozoic (ca. 1.2 Ga) pCO<sub>2</sub> levels to have been ≤0.36% (~10 PAL). This value supports results from Fe-silicate equilibria models and models based on constitutive mass balance of paleosols, which indicate pCO<sub>2</sub> levels of <5–25 PAL from 3.2 to 1.0 Ga (Hessler et al., 2004; Sheldon, 2006), which is within the range of Phanerozoic pCO<sub>2</sub> values derived from the GEOCARB III model (Berner and Kothavala, 2001).

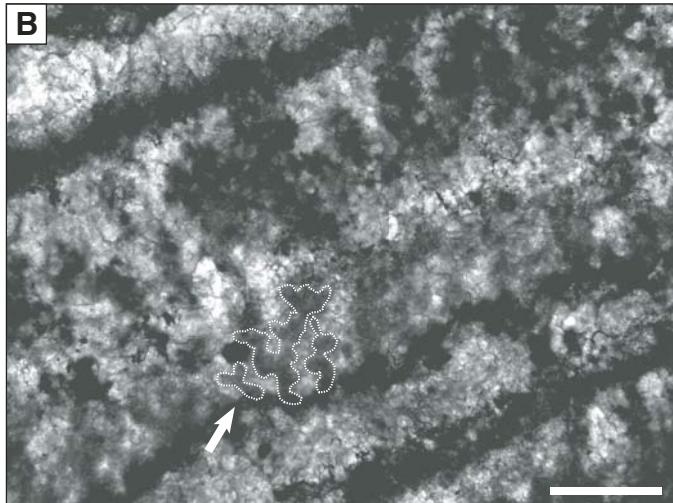
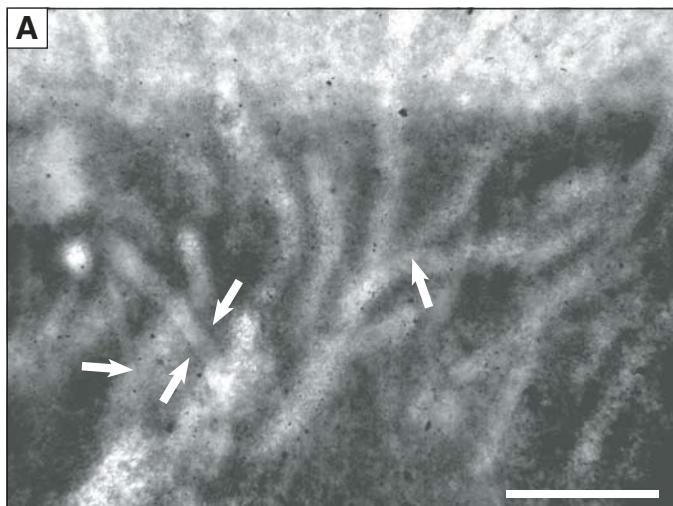
## MICROBIAL FABRICS OF THE SOCIETY CLIFFS FORMATION

The ca. 1.2 Ga Society Cliffs Formation (Kah et al., 2001), northernmost Baffin and Bylot Islands, Arctic Canada, consists of >600 m of peritidal dolostone deposited on a stable platform. Eastern and northeastern portions of the basin represent sedimentary deposition in intertidal to supratidal, low-energy, evaporative tidal-flat environments. Tidal-flat environments are characterized by low-diversity microbial populations that occur in discrete layers within low-relief mounds (<1 m in diameter), interlaminated with thin accumulations of detrital micrite and seafloor precipitates that encrust topographic surfaces. Penecontemporaneous silicification results in excellent preservation of primary microbial and precipitate fabrics and facilitates interpretation of otherwise dolomitized microfabrics (Kah and Knoll, 1996). Silicified mat fabrics consist primarily of coccoidal cyanobacteria (*Eoentophysalis* sp.) or monotypic filament mats (*Siphonophycus* sp.) that construct vertically oriented tufts, <0.5 mm to >10 mm in height, that are preserved through a combination of early lithification, burial by detrital micrite, and encrustation by carbonate precipitates (Kah and Knoll, 1996).

In addition to silicified mat fabrics, the Society Cliffs Formation locally preserves calcified microbial filaments and shrubs (Fig. 1), which occur interlaminated with seafloor precipitates and detrital micrite and adjacent to silicified mats. Calcified filaments occur as subvertical unbranched curved tubes, 15 µm wide, in dense micrite. The tubes have relatively constant diameter, and uniform thickness micritic walls are locally visible where filaments cross (Fig. 1A). Uniformity of wall thickness and absence of filament degradation, which would occur during post-mortem calcification, indicate sheath impregnation produced by *in vivo* calcification. *In vivo* cyanobacterial calcification is localized in the protective mucilaginous sheath surrounding cyanobacterial cells (Pentecost and Riding, 1986; Merz-Preiß, 2000). The size and shape of Society Cliffs calcified filaments closely resemble *Girvanella*, which has present-day analogs in *in vivo* CaCO<sub>3</sub>-impregnated sheaths of the scytonematacean cyanobacterium *Plectonema* (Riding, 1977).

Microbial shrubs are composed of fine microspar; they are 200–600 µm high and up to 200 µm wide and have irregular margins (Fig. 1B). They generally broaden upward and are digitate in their upper regions. On

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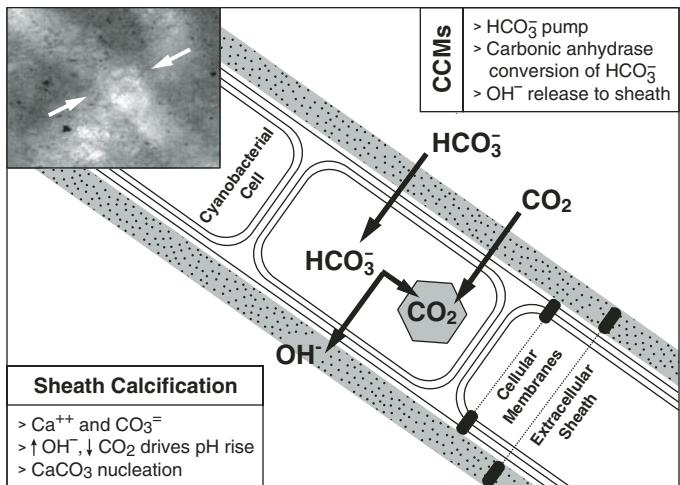


**Figure 1.** Society Cliffs Formation calcified cyanobacteria. **A:** Filament molds are 15  $\mu\text{m}$  in diameter and up to 1.2 mm in length. Uniform thickness walls (arrows) are typical of in vivo sheath calcification. Scale bar is 125  $\mu\text{m}$ . **B:** Calcified shrubs composed of dolomicrospar are ~200  $\mu\text{m}$  wide, 200–600  $\mu\text{m}$  high, and have irregular digitate margins (outlined). Structures of similar size, spacing, and orientation preserved in early diagenetic chert consist of vertically oriented microbial filament tufts. Scale bar is 500  $\mu\text{m}$ .

curved stromatolite surfaces, consistent vertical orientation of the shrubs suggests a phototactic growth response. In size and morphology, shrubs closely resemble *Angusticellularia* (= *Angulocellulararia*), a modern oscillatoriacean cyanobacteria that calcifies by micritic impregnation of thick irregular sheaths (Riding and Voronova, 1982). A filamentous origin for the Society Cliffs shrubs is further supported by the presence of filamentous tufts of *Siphonophycus* sp. in silicified regions adjacent to shrub carbonate.

#### CYANOBACTERIAL CALCIFICATION AND PALEO-PCO<sub>2</sub> ESTIMATES

Cyanobacterial calcification is primarily dependent on ambient carbonate saturation state (Kempe and Kazmierczak, 1994) and on pH changes in the microbial sheath resulting from photosynthetic uptake of CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> (Golubic, 1973; Arp et al., 2001). This latter effect is greatly enhanced by carbon concentrating mechanisms (CCMs), which include active HCO<sub>3</sub><sup>-</sup> transport into the cells, its conversion to CO<sub>2</sub>, and concomitant release of OH<sup>-</sup> ions that further raises sheath pH, promoting CaCO<sub>3</sub> nucleation (Fig. 2; Merz, 1992). Laboratory experiments show that



**Figure 2.** Inferred mechanism of in vivo calcification in response to carbon-concentrating mechanism (CCM) induction. Sheath is preserved via nucleation and impregnation by finely crystalline CaCO<sub>3</sub> as pH rises in response to OH<sup>-</sup> release due to active HCO<sub>3</sub><sup>-</sup> uptake (after Riding, 2006).

present-day cyanobacteria and algae induce CCMs in response to atmospheric CO<sub>2</sub> below 0.36% (Badger et al., 2002, and references therein). It is reasoned, therefore, that the inception of cyanobacterial sheath calcification in the Proterozoic reflects reduction of atmospheric CO<sub>2</sub> to this critical threshold (Riding, 2006).

Photosynthetic carbon uptake within robust and highly productive benthic mats can result in microenvironmental CO<sub>2</sub> concentrations well below equilibrium levels. In these environments, CCM induction could occur even when atmospheric CO<sub>2</sub> exceeds 0.36%. In the Society Cliffs examples, however, calcified cyanobacteria occur as thin, submillimeter layers that are overlain by micritic drapes or precipitates that encrust microbial topography, suggesting that tufts maintained contact with seawater throughout growth. Under these circumstances, we infer that CCM induction occurred under equilibrium conditions, which indicates atmospheric CO<sub>2</sub> levels close to or below 0.36%. Prior to discovery of Society Cliffs calcified cyanobacteria, the oldest reports of in vivo sheath calcification included *Girvanella* in the 750–700 Ma Draken Group of Spitzbergen (Knoll et al., 1993) and similar structures in the ca. 800 Ma Little Dal Group of northwest Canada (Turner et al., 1993). Calcified cyanobacteria in the Society Cliffs Formation predate these Neoproterozoic occurrences by at least 400 m.y.

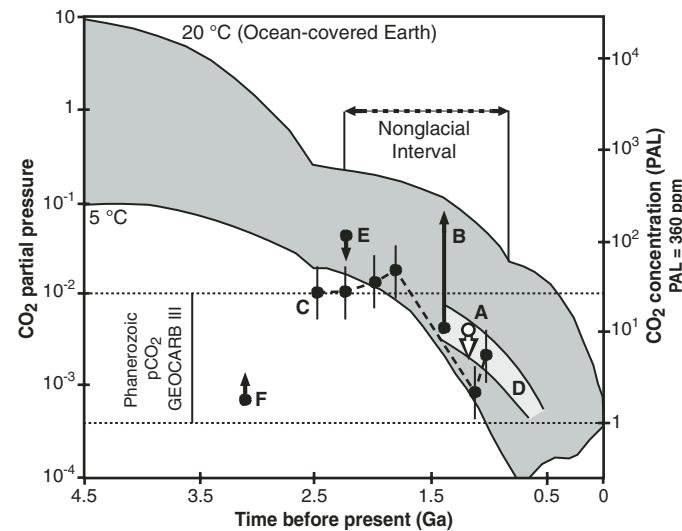
#### DISCUSSION

The presence of calcified cyanobacteria in the Society Cliffs Formation indicates atmospheric pCO<sub>2</sub> levels  $\leq$  0.36% (~10 PAL) at 1.2 Ga. Assuming solar luminosity 90% of the present-day levels in the late Mesoproterozoic (Gough, 1981), a one-dimensional climate model (Kasting, 1987) indicates that CO<sub>2</sub> concentrations of ~10 PAL would result in average global surface temperatures of ~7 °C. However, the absence of convincing glacial deposits (Williams and Schmidt, 1996; Young, 1998) suggests that Mesoproterozoic temperatures were likely closer to 15–20 °C (Kasting, 1987), thereby requiring the presence of additional greenhouse gases. Climate models based on addition of methane to an atmosphere with present-day CO<sub>2</sub> levels show that a 100 ppm increase in methane would result in an increase in global surface temperature of <12° (Pavlov et al., 2003). Thus, we suggest that methane concentrations in the late Mesoproterozoic may have reached 100–200 ppm, which would have required at least a tenfold increase in methane flux.

Sustaining elevated methane is problematic in an oxygenated Earth system that favors both lower methane production via competition of sulfate-reducing bacteria with methanogens for organic matter and increased removal of methane via chemical oxidation, consumption by aerobic methanotrophs (Ren et al., 1997), and anaerobic decomposition (Boetius et al., 2000). Alternatively, biospheric oxygen, and consequently marine sulfate concentrations (Habicht et al., 2002), may have remained low throughout much of the Proterozoic. Based on microbial physiology, Canfield and Teske (1996) proposed that disproportionating sulfur-oxidizing bacteria would not have evolved until  $pO_2$  reached 5%–15% PAL. Discrimination of sulfur disproportionation via  $\Delta^{33}S$  analysis suggests that these levels were first reached at ca. 1.3 Ga (Johnston et al., 2005), which supports arguments that the relatively restricted S-isotope range observed in pyrite prior to ca. 750 Ma likely resulted from marine sulfate concentrations so low as to limit the isotopic expression of bacterial sulfur cycling (Kah et al., 2001). Rate-dependent modeling of S-isotope change recorded in carbonate-associated sulfate further suggests that marine sulfate concentrations remained <15% of modern values until at least 1.2 Ga and may not have risen substantially until the mid-Neoproterozoic (Kah et al., 2004). Together, these lines of reasoning support low atmospheric oxygen and potentially elevated methane levels throughout the Mesoproterozoic.

These paleo- $pCO_2$  estimates of  $\leq 10$  PAL for the ca. 1.2 Ga Society Cliffs calcified cyanobacteria (Fig. 2) are considerably lower than >10–200 PAL estimates based on the isotopic composition of ca. 1.4 Ga acritarchs (Kaufman and Xiao, 2003). Nonetheless, Society Cliffs values are similar to those obtained by constitutive mass balance calculations (Sheldon, 2006) of ca. 1.1 Ga Sturgeon Falls (Zbinden et al., 1988) and 0.98 Ga Sheigra (Retallack and Mindszenty, 1994) paleosols (Fig. 3). Society Cliffs  $pCO_2$  estimates are also consistent with both textural evidence from marine carbonates and model calculations that suggest a decrease in the size of the marine dissolved inorganic carbon (DIC) reservoir through the Mesoproterozoic (Bartley and Kah, 2004). In this model, DIC reservoir size affects the sensitivity of the marine carbon isotopic system to biogeochemical perturbation: a DIC reservoir >10× present effectively buffers the ocean against isotopic change, whereas a DIC reservoir <10× present provides an insufficient buffer against isotopic change. By calculating DIC reservoir size from rates of isotopic response and recovery, Kah and Bartley (2004) suggested that marine DIC, and thus atmospheric  $pCO_2$ , was likely to be >10× present prior to ca. 1.3 Ga, 7–10× present in the late Mesoproterozoic, and 2× present in the latest Neoproterozoic (Fig. 3). Prior to ca. 1.3 Ga, elevated DIC resulted in limited isotopic change, which hinders utilization of the C-isotopic record for accurate determination of DIC reservoir size. Nonetheless, geologically rapid isotopic shifts in the latest Paleoproterozoic (Melezik et al., 1999) suggest that Archean marine DIC may not have significantly exceeded 10× present, and may have been as low as 2× present (Hessler et al., 2004).

Emerging views of the Proterozoic global carbon cycle suggest that atmospheric  $pCO_2$  may not have differed substantially from that estimated for the Phanerozoic (Fig. 3). Because in vivo cyanobacterial calcification is significantly influenced by  $pCO_2$ -driven changes in carbonate saturation state (Riding, 2006), we suggest calcified cyanobacteria may have been more widespread in the Precambrian than currently recognized and may have played a critical role in constraining trends in  $pCO_2$ . For example, current estimates suggest that  $pCO_2$  may have been >10 PAL at 1.4 Ga, but they do not constrain maximum  $pCO_2$  (Kaufman and Xiao, 2003; Bartley and Kah, 2004). Discovery of sheath calcified cyanobacteria of this age would constrain  $pCO_2$  to  $\leq 10$  PAL. Similarly, elevated  $pCO_2$  estimates for the Paleoproterozoic (Rye et al., 1995; Sheldon, 2006) suggest that cyanobacterial sheath calcification may have been absent at this time, whereas lower  $pCO_2$  estimates for the Archean (Hessler et al., 2004) would have permitted CCM-induced sheath calcification.



**Figure 3. Estimates of Proterozoic  $pCO_2$ .** Shaded region represents model  $pCO_2$  estimates from Kasting (1987), wherein upper and lower boundaries reflect average surface temperatures for an ice-free ( $20^\circ\text{C}$ ) and ice-covered ( $5^\circ\text{C}$ ) Earth. Limits of Phanerozoic  $pCO_2$  from GEOCARB III model (Berner and Kothavala, 2001) are denoted by dashed lines. Cyanobacterial calcification (A; present study) indicates Mesoproterozoic (1.6–1.0 Ga)  $pCO_2$  <10 present atmospheric level (PAL), similar to minimum estimates based on acritarch isotopic composition (B; Kaufman and Xiao, 2003), estimates derived from paleosol mass balances (C; Sheldon, 2006), and model estimates derived from C-isotope reservoir modeling (D; Kah and Bartley, 2004). These estimates, together with those for Paleoproterozoic (2.5–1.6 Ga) (E; Rye et al., 1995) and Archean (3.8–2.5 Ga) (F; Hessler et al., 2004), suggest that  $pCO_2$  may never have been substantially higher than Phanerozoic levels, which would have required additional greenhouse gases to compensate for decreased solar luminosity, particularly during ice-free intervals.

## CONCLUSIONS

The induction of  $CO_2$ -concentrating mechanisms during photosynthesis, and resultant in vivo calcification of cyanobacterial sheaths, provides an ecophysiological link to atmospheric  $pCO_2$ . Calcified sheaths from the Society Cliffs Formation represent the first known occurrence of in vivo calcification, and they indicate  $pCO_2$  levels  $\leq 10$  PAL in the late Mesoproterozoic. Estimates are similar to those inferred from Mesoproterozoic paleosols and are consistent with estimates of 7–10 PAL from time-dependent models of marine carbon isotope change. Relatively low concentrations of atmospheric  $CO_2$  would require addition of other greenhouses gases, e.g., 100–200 ppm methane, to counteract the effects of lower solar luminosity on Earth surface temperature. Such atmospheric methane concentrations are consistent with estimates of low  $pO_2$  and marine sulfate concentration that reflect protracted oxygenation of the Proterozoic biosphere. The potential importance of methane on early Earth requires critical evaluation of paleo- $pCO_2$ . Recent models (Fig. 3) suggest that  $pCO_2$  in the Precambrian may not have differed significantly from Phanerozoic values, yet evidence for cyanobacterial calcification has not been confidently recognized in the Paleoproterozoic or Archean. Because in vivo cyanobacterial calcification provides an empirical indication of  $pCO_2$  levels  $\leq 10$  PAL, discovery of these distinctive microfossils in rocks older than 1.2 Ga could provide critical constraints on the evolution of Earth's early atmosphere.

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## REFERENCES CITED

- Arp, G., Reimer, A., and Reitner, J., 2001, Photosynthesis-induced biofilm calcification and calcium concentrations in Phanerozoic oceans: *Science*, v. 292, p. 1701–1704, doi: 10.1126/science.1057204.
- Badger, M.R., Hanson, D., and Price, G.D., 2002, Evolution and diversity of CO<sub>2</sub> concentrating mechanisms in cyanobacteria: *Functional Plant Biology*, v. 29, p. 161–173, doi: 10.1071/PP01213.
- Bartley, J.K., and Kah, L.C., 2004, Marine carbon reservoir, C<sub>org</sub>-C<sub>carb</sub> coupling, and the evolution of the Proterozoic carbon cycle: *Geology*, v. 32, p. 129–132, doi: 10.1130/G19939.1.
- Berner, R.A., and Kothavala, Z., 2001, GEOCARB III: A revised model of atmospheric CO<sub>2</sub> over Phanerozoic time: *American Journal of Science*, v. 301, p. 182–204, doi: 10.2475/ajs.301.2.182.
- Boetius, A., Ravenschlag, K., Schubert, C.J., Rickert, D., Widdel, F., Gieseke, A., Amann, R., Jørgensen, B.B., Witte, U., and Pfannkuche, O., 2000, A marine microbial consortium apparently mediating anaerobic oxidation of methane: *Nature*, v. 407, p. 623–626, doi: 10.1038/35036572.
- Canfield, D.E., 1998, A new model for Proterozoic ocean chemistry: *Nature*, v. 396, p. 450–453, doi: 10.1038/24839.
- Canfield, D.E., and Teske, A., 1996, Late Proterozoic rise in atmospheric oxygen inferred from phylogenetic and sulphur-isotope studies: *Nature*, v. 382, p. 127–132, doi: 10.1038/382127a0.
- Farquhar, J., Bao, H., and Thiemens, M., 2000, Atmospheric influences of the Earth's earliest sulfur cycle: *Science*, v. 289, p. 756–758, doi: 10.1126/science.289.5480.756.
- Goldblatt, C., Lenton, T.M., and Watson, A.J., 2006, Bistability of atmospheric oxygen and the great oxidation: *Nature*, v. 443, p. 683–686, doi: 10.1038/nature05169.
- Golubic, S., 1973, The relationship between blue-green algae and carbonate deposits, in Carr, N., and Whitton, B.A., eds., *The Biology of Blue-Green Algae*: Oxford, Blackwell, p. 434–472.
- Gough, D.O., 1981, Solar interior structure and luminosity variations: *Solar Physics*, v. 74, p. 21–34, doi: 10.1007/BF00151270.
- Habicht, K.S., Gade, M., Thamdrup, B., Berg, P., and Canfield, D.E., 2002, Calibration of sulfate levels in the Archean Ocean: *Science*, v. 298, p. 2372–2374, doi: 10.1126/science.1078265.
- Hessler, A.M., Lowe, D.R., Jones, R.L., and Bird, D.K., 2004, A lower limit for atmospheric carbon dioxide levels 3.2 billion years ago: *Nature*, v. 428, p. 736–738, doi: 10.1038/nature02471.
- Johnston, D.T., Wing, B.A., Farquhar, J., Kaufman, A.J., Strauss, H., Lyons, T.W., Kah, L.C., and Canfield, D.E., 2005, Active microbial sulphur disproportionation in the Mesoproterozoic: *Science*, v. 310, p. 1477–1479, doi: 10.1126/science.1117824.
- Kah, L.C., and Bartley, J.K., 2004, Effect of marine carbon reservoir size on the duration of carbon isotope excursions: Interpreting the Mesoproterozoic carbon isotope record: *Geological Society of America Abstracts with Programs*, v. 36, no. 5, p. 78.
- Kah, L.C., and Knoll, A.H., 1996, Microbenthic distribution of Proterozoic tidal flats: Environmental and taphonomic considerations: *Geology*, v. 24, p. 79–82, doi: 10.1130/0091-7613(1996)024<0079:MDOPTF>2.3.CO;2.
- Kah, L.C., Lyons, T.W., and Chesley, J.T., 2001, Geochemistry of a 1.2 Ga carbonate-evaporite succession, northern Baffin and Blyot Islands: Implications for Mesoproterozoic marine evolution: *Precambrian Research*, v. 111, p. 203–234, doi: 10.1016/S0301-9268(01)00161-9.
- Kah, L.C., Lyons, T.W., and Frank, T.D., 2004, Low marine sulphate and protracted oxygenation of the Proterozoic biosphere: *Nature*, v. 431, p. 834–838, doi: 10.1038/nature02974.
- Kasting, J.F., 1987, Theoretical constraints on oxygen and carbon dioxide concentrations in the Precambrian atmosphere: *Precambrian Research*, v. 34, p. 205–229, doi: 10.1016/0301-9268(87)90001-5.
- Kasting, J.F., 2005, Methane and climate during the Precambrian era: *Precambrian Research*, v. 137, p. 119–129, doi: 10.1016/j.precamres.2005.03.002.
- Kasting, J.F., Zahnle, K.J., and Walker, J.C.G., 1983, Photochemistry of methane in the Earth's early atmosphere: *Precambrian Research*, v. 20, p. 121–148, doi: 10.1016/0301-9268(83)90069-4.
- Kaufman, A.J., and Xiao, S., 2003, High CO<sub>2</sub> levels in the Proterozoic atmosphere estimated from analyses of individual microfossils: *Nature*, v. 425, p. 279–282, doi: 10.1038/nature01902.
- Kempe, S., and Kazmierczak, J., 1994, The role of alkalinity in the evolution of ocean chemistry, organization of living systems, and biocalcification processes: *Bulletin de l'Institut Océanographique (Monaco)*, v. 13, p. 61–117.
- Knoll, A.H., Fairchild, I.J., and Swett, K., 1993, Calcified microbes in Neoproterozoic carbonates: implications for our understanding of the Proterozoic/Cambrian transition: *Palaios*, v. 8, p. 512–525.
- Melezhik, V.A., Fallick, A.E., Medvedev, P.V., and Makarikhin, V.V., 1999, Extreme <sup>13</sup>C enrichment in ca. 2.0 Ga magnesite-stromatolites-dolomite “red beds” association in a global context: A case for the world-wide signal enhanced by a local environment: *Earth-Science Reviews*, v. 48, p. 71–120, doi: 10.1016/S0012-8252(99)00044-6.
- Merz, M.U.E., 1992, The biology of carbonate precipitation by cyanobacteria: *Facies*, v. 26, p. 81–102, doi: 10.1007/BF02539795.
- Merz-Preiß, M., 2000, Calcification in cyanobacteria, in Riding, R., and Awramik, S.M., eds., *Microbial Sediments*: Berlin, Springer-Verlag, p. 50–56.
- Owen, T., Cess, R.D., and Ramanathan, V., 1979, An enhanced carbon dioxide greenhouse to compensate for reduced solar luminosity: *Nature*, v. 277, p. 640–642, doi: 10.1038/277640a0.
- Pavlov, A.A., Kasting, J.F., and Brown, L.L., 2001, UV-shielding of NH<sub>3</sub> and O<sub>2</sub> by organic hazes in the Archean atmosphere of early Earth: *Journal of Geophysical Research*, v. 106, p. 23,267–23,287, doi: 10.1029/2000JE001448.
- Pavlov, A.A., Hurtgen, M.T., Kasting, J.F., and Arthur, M.A., 2003, Methane-rich Proterozoic atmosphere?: *Geology*, v. 31, p. 87–90, doi: 10.1130/0091-7613(2003)031<0087:MRPA>2.0.CO;2.
- Pentecost, A., and Riding, R., 1986, Calcification in cyanobacteria, in Leadbeater, B.S.C., and Riding, R., eds., *Biomineralization in Lower Plants and Animals: Systematics Association Special Volume 30*: Oxford, Clarendon Press, p. 73–90.
- Ren, T., Amaral, J.A., and Knowles, R., 1997, The response of methane consumption by pure cultures of methanotrophic bacteria to oxygen: *Canadian Journal of Microbiology*, v. 43, p. 925–928.
- Retallack, G.J., and Mindszenty, A., 1994, Well preserved Precambrian paleosols from northwest Scotland: *Journal of Sedimentary Research*, v. A64, p. 264–281.
- Riding, R., 1977, Calcified *Plectonema* (blue-green algae), a recent example of *Girvanella* from Aldabra Atoll: *Palaeontology*, v. 20, p. 33–46.
- Riding, R., 2006, Cyanobacterial calcification, carbon dioxide concentrating mechanisms, and Proterozoic-Cambrian changes in atmospheric composition: *Geobiology*, v. 4, p. 299–316, doi: 10.1111/j.1472-4669.2006.00087.x.
- Riding, R., and Voronova, L., 1982, Affinity of the Cambrian alga *Tubomorphophyton* and its significance for the Epiphytaceae: *Palaeontology*, v. 25, p. 869–878.
- Rye, R., Kuo, P.H., and Holland, H.D., 1995, Atmospheric carbon-dioxide concentrations before 2.2-billion years ago: *Nature*, v. 378, p. 603–605, doi: 10.1038/378603a0.
- Sagan, C., and Mullen, G., 1972, Earth and Mars: Evolution of atmospheres and surface temperatures: *Science*, v. 177, p. 52–56, doi: 10.1126/science.177.4043.52.
- Schrag, D.P., Berner, R.A., Hoffman, P.F., and Halverson, G.P., 2002, On the initiation of a snowball Earth: *Geochemistry, Geophysics, Geosystems*, v. 3, doi: 10.1029/2001GC000219.
- Sheldon, N.D., 2006, Precambrian paleosols and atmospheric CO<sub>2</sub> levels: *Precambrian Research*, v. 147, p. 148–155, doi: 10.1016/j.precamres.2006.02.004.
- Turner, E.C., Narbonne, G.M., and James, N.P., 1993, Neoproterozoic reef microstructures from the Little Dal Group, northwestern Canada: *Geology*, v. 21, p. 259–262, doi: 10.1130/0091-7613(1993)021<0259:NRMFTL>2.3.CO;2.
- Williams, G.E., and Schmidt, P.W., 1996, Origin and palaeomagnetism of the Mesoproterozoic Gangau tilloid (basal Vindhyan Supergroup), central India: *Precambrian Research*, v. 79, p. 307–325, doi: 10.1016/0301-9268(95)00101-8.
- Young, G.M., 1998, Non-glacial origin for conglomerates and associated rocks at the base of the Mesoproterozoic (ca. 1.2 Ga) Stoer Group (Torridonian) of northwest Scotland: *Geological Society of America Abstracts with Programs*, v. 30, no. 7, p. 291.
- Zbinden, E.A., Holland, H.D., Feakes, C.R., and Dobos, S.K., 1988, The Sturgeon Falls paleosol and the composition of the atmosphere 1.1 Ga BP: *Precambrian Research*, v. 42, p. 141–163, doi: 10.1016/0301-9268(88)90014-9.

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